

METHODS FOR MODULATING IKK α ACTIVITY

CROSS-REFERENCE TO RELATED APPLICATIONS

5 This patent application claims benefit to US provisional patent application No. 60/431,825, filed December 9, 2002.

BACKGROUND OF THE INVENTION

10 This invention relates to the field of, inflammatory diseases and autoimmune diseases and the treatment thereof through the modulation of IKK α activity and the modulation of genes under the control of IKK α .

BACKGROUND INFORMATION

15 The NF- κ B or nuclear factor κ B is a transcription factor that plays a critical role in inflammatory diseases by inducing the expression of a large number of proinflammatory and anti-apoptotic genes. These include cytokines such as IL-1, IL-2, IL-11, TNF- α and IL-6, chemokines including IL-8, GRO1 and RANTES, as well as other proinflammatory molecules including COX-2 and cell adhesion molecules such as ICAM-1, VCAM-1, and E-selectin. Pahl HL, (1999) *Oncogene* 18, 6853-6866; Jobin et al, (2000) *Am. J. Physiol. Cell. Physiol.* 278: 451-462. Under resting conditions, NF- κ B is present in the cytosol of cells as a complex with I κ B. The I κ B family of proteins serve as inhibitors of NF- κ B, interfering with the function of its nuclear localization signal (see for example U. Siebenlist et al, (1994) *Ann. Rev. Cell Bio.*, 10: 405). Upon disruption of the I κ B- NF- κ B complex following cell activation, NF- κ B translocates to the nucleus and activates gene
25 transcription. Disruption of the I κ B- NF- κ B complex and subsequent activation of NF- κ B is initiated by degradation of I κ B.

 Activators of NF- κ B mediate the site-specific phosphorylation of two amino terminal serines in each I κ B which makes nearby lysines targets for ubiquitination, thereby
30 resulting in I κ B proteasomal destruction. NF- κ B is then free to translocate to the nucleus and bind DNA leading to the activation of a host of inflammatory response target genes.

Baldwin, A., Jr., (1996) *Annu Rev Immunol* 14: 649-683, Ghosh, S. et al, (1998) *Annu Rev Immunol* 16, 225-260. Recent evidence has shown that NF- κ B subunits dynamically shuttle between the cytoplasm and the nucleus but a dominant acting nuclear export signal in I κ B α ensures their transport back to the cytoplasm.

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Even though NF- κ B is largely considered to be a transcriptional activator, under certain circumstances it can also be involved in directly repressing gene expression (reviewed in Baldwin, A., Jr., (1996) *Annu. Rev. Immunol.*, 14: 649-683; Ghosh, S. et al. (1998) *Annu. Rev. Immunol.*, 16: 225-260).

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The phosphorylation of I κ B is a major triggering event in regulation of the NF- κ B pathway. Since the abnormal regulation of the NF- κ B pathway is known to correlate with inflammatory disease, the regulation of I κ B phosphorylation is understood as an important area for disease intervention. The search for the kinase responsible for the inducible phosphorylation of I κ B has been one of the major focuses in the NF- κ B field. I κ B phosphorylation is mediated by a high molecular weight signalsome complex consisting of at least three components: two I κ B kinases IKK α , IKK β and a non-catalytic regulatory subunit NEMO (reviewed in Mercurio, F. et al, (1999) *Oncogene*, 18: 6163-6171; Barkett, M. et al, (1999) *Oncogene*, 18: 6910-6924; Karin, M., (1999) *Oncogene*, 18: 867-6874). A great deal of study has been performed to determine the respective roles that each of the components play in the regulation of NF- κ B with the belief that a greater understanding of the roles might lead to the development of new methods and approaches for the treatment of inflammatory diseases. Two molecules of NEMO are believed to orchestrate the assembly of the IKKs into the high molecular weight signalsome complex at least in part by binding to specific carboxy-terminally conserved residues of both IKK α and IKK β termed the NEMO binding domain or NBD. Krappmann, D. et al, (2000) *J. Biol Chem* 275: 29779-29787; Li, X.H. et al, (2001) *J. Biol. Chem.*, 276: 4494-4500; Hatada, E.N. et al, (2000) *Current Opinion in Immunology*, 12: 52-58; May, M. J. et al, (2000) *Science*, 289: 1550-1554. NEMO may also facilitate the recruitment of I κ B α to the IKK complex. Yamamoto, Y. et al, (2001) *J. Biol. Chem.*, 276: 36327-36336. The two catalytic IKK subunits

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differentially respond via NEMO to an array of signal induced, upstream kinase activities culminating in the coordinated phosphorylation of a pair of serines in their MAPK-like T activation loops by an unknown mechanism.

5 The roles of the IKKs in NF- κ B activation were studied in mice lacking IKK β , IKK α or NEMO. Li, Q. et al, (1999) *Science*, 28: 321-325; Li, Z. et al (1999) *J. Exp. Med.* 189: 1839-1845; Tanaka, M. et al, (1999) *Immunity*, 10: 421-429; Li, Q. et al, (1999) *Genes Dev.*, 13: 1322-1328; Hu, Y. et al. (1999) *Science*, 284: 316-320; Takeda, K. et al, (1999) *Science*, 284: 313-316. Akin to mice genetically deficient for the NF- κ B p65
10 subunit (Beg, A. A. et al, (1995) *Nature*, 376: 167-170), murine embryos genetically null for either IKK β or NEMO succumbed to severe liver apoptosis in utero due to a virtually complete block in NF- κ B activation. Li, Q. et al, (1999) *Science*, 284: 321-325; Li, Z. et al. (1999) *J. Exp. Med.*, 189: 1839-1845; Tanaka, M. et al, (1999) *Immunity*, 10: 421-429; Rudolph, D. et al, (2000) *Genes and Dev.* 14: 854-862; Schmidt-Supprian, M. et
15 al, (2000) *Mol. Cell*, 5: 981-992; Makris, C. et al, (2000) *Mol. Cell*, 5: 969-979. These IKK β and NEMO KO animals were severely if not completely deficient for both cytokine mediated I κ B degradation and nuclear NF- κ B DNA binding activity. Li, Q. et al, (1999) *Science*, 284: 321-325; Li, Z. et al, (1999) *J. Exp. Med.* 189: 1839-1845; Tanaka, M. et al, (1999) *Immunity*, 10: 421-429; Rudolph, D. et al. (2000) *Genes and Dev.*, 14: 854-862;
20 Schmidt-Supprian, M. et al, (2000) *Mol. Cell*, 5: 981-992; Makris, C. et al, (2000) *Mol. Cell* 5: 969-979.

 In contrast, to the IKK β and NEMO KO mice, IKK α null animals died perinatally due to severe skin, limb and skeletal abnormalities caused by a block in the terminal
25 differentiation of epidermal keratinocytes. Li, Q. et al, (1999) *Genes Dev.* 13: 1322-1328; Hu, Y. et al, (1999) *Science*, 284: 316-320; Takeda, K. et al. (1999) *Science*, 284: 313-316. Subsequent work revealed that IKK α , (independent of both its kinase activity and NF- κ B), controls the production of a soluble factor that induces keratinocyte differentiation. Hu, Y., Baud, V. et al, (2001) *Nature*, 410: 710-714. Furthermore, IKK α null embryos appeared to
30 be phenotypically normal for both cytokine induced I κ B α degradation, NF- κ B nuclear

translocation and NF- κ B DNA binding activity. Hu, Y. et al, (1999) *Science*, 284: 316-320; Takeda, K. et al, (1999) *Science*, 284: 313-316. In addition, an independent study in cultured mammalian cells employing transfection conditions that avoided over-expression artifacts concluded that the cytokine controlled activation of NF- κ B induction was an *in vivo* function of IKK β and not IKK α . Delhase, M. et al, (1999) *Science*, 284: 309-313.

This body of work has led to the well-accepted belief in the art that IKK β alone is essential for NF- κ B activation by inflammatory response mediators. Karin, M. (1999) *Oncogene*, 18: 6867-6874; Hatada, E.N. et al, (2000) *Current Opinion in Immunology*, 12: 52-58; Karin, M. et al. (2000) *Annu. Rev. Immunol.*, 18: 621-663. More recently and in keeping with its separate and distinct functions from IKK β , IKK α has been shown to possess at least two additional novel *in vivo* functions: (a) it is essential for B lymphocyte maturation (Kaisho, T. et al, (2001) *J. Exp. Med.* 193: 417-426) and Peyers patch formation via an LT β R and NIK dependent signaling pathway (Matsushima, A. et al, (2001) *J. Exp. Med.* 193: 631-636), wherein it is required to target the cytokine induced processing of the NF- κ B2 (p100) precursor to produce the functional NF- κ B p52 subunit (Senftleben, U. et al, (2001) *Science*, 293: 1495-1499) and (b) it is required for the proliferation of mammary epithelial cells in response to RANK ligand but not TNF α signaling to activate cyclin D1. Cao, Y., Bonizzi, G. et al, (2001) *Cell*, 107: 763-775. Independent of these studies, IKK β was reported to phosphorylate an I κ B-like destruction motif in p50's p105 precursor, which produces a recognition site for β TrCP-containing SCF ubiquitin ligases with subsequent polyubiquitination of p105 causing its complete proteasomal destruction and the induced release of DNA binding p50 homodimers (Heissmeyer, V. et al, (1999) *Embo. J.*, 18: 4766-4778; Heissmeyer, V. et al, (2001) *Mol. Cell Biol.*, 21: 1024-1035), providing additional support for the notion that IKK β and IKK α have distinct roles in NF- κ B activation.

In addition to the well accepted belief of induced nuclear translocation of NF- κ B dependent gene expression, an alternative mechanism has emerged that involves the phosphorylation of the p65 transactivation subunit. The protein kinase A catalytic subunit phosphorylates p65 which leads to the association of p65 and the p300 transcriptional

coactivator. Zhong, H. et al, *Mol. Cell.*, (1998) 1: 661-671. Cells from GSK3 and T2K knockout mice are capable of inducing NF- κ B nuclear translocation but are deficient in stimulating transactivation functions of NF- κ B. Hoeflich et al, (2000) *Nature*, 406: 86-90; Bonnard, M. et al, (2000) *Embo. J.*, 19: 4976-4985. Thus, NF- κ B dependent gene
5 transcription is regulated at other step(s) in addition to I κ B α degradation and NF- κ B translocation.

Recently, it has been shown that in mouse embryonic fibroblasts, IKK α is required for NF- κ B-mediated gene transcription in response to proinflammatory cytokine TNF α and IL-1 β . Li, X. et al, (2002) *J. Biol. Chem.*, 277: 45129-45140. This is in dramatic
10 contrast to the generally accepted view of IKK α being dispensable for TNF and IL-1 induced gene transcription mediated by NF- κ B. Hu, Y. et al, (1999) *Science*, 284: 316-320; Takeda, K. et al, (1999) *Science*, 284: 313-316. One way to demonstrate that IKK α is also needed for TNF α - induced NF- κ B dependent gene transcription in human cells, is to
15 specifically knock down the expression or activity of IKK α .

The function of a gene can be determined on the basis of the behavior of cells in which the level of gene expression or level of activity of the gene product has been reduced. Experimental procedures can be used to specifically inactivate or silence a target
20 gene or inhibit the activity of its gene product. Inhibition of protein activity can be brought about at the level of gene transcription, protein translation or post translational modifications. For instance, the activity of a protein can be inhibited by directly inhibiting the activity of the protein such as altering a catalytic domain or alternatively by reducing the amount of the protein in the cell by reducing the amount of mRNA encoding the
25 protein. In each case the level of protein activity in the cell is reduced. Various techniques can be used to knock down the activity of a protein and these include knockout technologies (antibodies, antisense RNA, and RNA interference) and compounds that specifically inhibit the protein activity. Antisense RNAs directed to IKK α has been reported for use in the inhibition of IKK α expression. US Patent No. 6,395,545.

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It is anticipated that compounds capable of modulating the expression of IKK α , and/or modulating the activity of IKK α may provide for a novel class of agents with activity toward a variety of inflammatory and autoimmune diseases such as osteoarthritis, reperfusion injury, asthma, multiple sclerosis, Guillain-Barre syndrome, Crohn's disease, ulcerative colitis, psoriasis, graft versus host disease, systemic lupus erythematosus, rheumatoid arthritis, Alzheimer's disease, toxic shock syndrome, inflammatory bowel disease, insulin-dependent diabetes mellitus, acute and chronic pain as well as symptoms of inflammation and cardiovascular disease, stroke, myocardial infarction alone or following thrombolytic therapy, thermal injury, adult respiratory distress syndrome (ARDS), multiple organ injury secondary to trauma, acute glomerulonephritis, dermatoses with acute inflammatory components, acute purulent meningitis or other central nervous system disorders, Grave's disease, myasthenia gravis, scleroderma and atopic dermatitis.

RNA interference (RNAi) is a technique that can be used to knockdown the activity of genes and their protein products in a specific manner. RNAi was first used in the Nematode worm *Caenorhabditis elegans* as a response to double stranded RNA (dsRNA) that resulted in the gene knockdown specific manner. Fire, A. et al, (1998) *Nature*, 391: 806-811. RNAi is a process whereby a double stranded RNA (dsRNA) of a sequence that is homologous to a target gene can be used to cause the degradation of messenger RNA (mRNA) transcribed from that target gene. Sharp, P.A., (2001) *Genes Dev.*, 15: 485-490. Initiation of gene silencing or gene inactivation occurs upon recognition of dsRNA by the cells machinery that convert the silencing trigger to 21-25 nucleotides RNAs. Hannon, (2002) *Nature*, 418: 244-250.

The mediators of sequence-specific messenger RNA degradation are 21- and 22-nucleotide small interfering RNAs (siRNAs) generated by ribonuclease III cleavage from longer dsRNAs. In vitro synthesized 21-nucleotide siRNA duplexes specifically suppress expression of endogenous and heterologous genes in different mammalian cell lines, including human embryonic kidney and HeLa cells. Elbashir S. et al, (2001) *Nature*, 411: 494-498. Therefore, 21-nucleotide siRNA duplexes provide a new tool for studying gene function in mammalian cells and may be used as gene-specific therapeutics. However,

effective gene silencing is only caused by a subset of siRNAs complementary to the mRNA target. McManus MT et al, (2002) *J. Immunol.* 169: 5754-60. Thus, design of multiple siRNA oligos and extensive testing are required to obtain a potent siRNA oligo. McManus MT et al, (2002) *J. Immunol.* 169: 5754-60.

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The ability to specifically knock down expression of a target gene by si RNA has many benefits. For example si RNA could be used to mimic true genetic knockout animals to study gene function. There have been reports of using siRNA for various purposes including the inhibition of luciferase gene expression in human cells, (see US
10 Patent Application No. 2002/0132788); HIV-1 Cellular receptor CD4 (Sharp et al, (2002) *Nature Medicine*, 8: 681-686); HIV accessory genes, vif and nef (Nature Advance Online Publication, June 26, 2002 (doi:10.1038/nature00896); HPV E6 and E7 gene expression. Jiang M., *Oncogene*, (2002), 21:6041-6048); Subtype- and species-specific knockdown of protein kinase C (Irie N. et al, *Biochem. Biophys. Res. Commun.*, (2002) 298: 738-743.

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BRIEF SUMMARY OF THE INVENTION

A first embodiment of the invention provides a method for modulating NF- κ B dependent gene transcription in a cell, said method comprised of modulating IKK α activity
20 in the cell.

A second embodiment of the invention provides for selectively modulating expression of a gene whose transcription is regulated by IKK α , the method comprising modulating IKK α activity such that expression of the gene is modulated.

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A third embodiment of the invention provides for modulating NF- κ B dependent gene transcription by administration of siRNA directed to IKK α to cells. SiRNA can be 21 to 25 nucleotides in length and hybridize to a nucleic acid molecule encoding human IKK α .

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A fourth embodiment of the invention provides a method for treating autoimmune and inflammatory disease in a mammal wherein the method is comprised of modulating IKK α expression or IKK α activity.

5 A fifth embodiment of the invention provides a method for treating autoimmune and inflammatory disease in a mammal wherein the method is comprised of modulating IKK α expression or IKK α activity by administration of siRNA directed to IKK α .

 A sixth embodiment of the invention provides a method for reducing the effects of
10 TNF α induced genes in cells, the method comprised of reducing IKK α activity in a cell.

 A seventh embodiment of the invention provides a method for reducing the effects of NF- κ B induced gene transcription, the method comprised of reducing IKK α activity in a cell.

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 An eighth embodiment of the invention provides a siRNA composition that when administered to a cell modulates IKK α activity.

 A ninth embodiment of the invention provides a method for identifying a
20 compound for the treatment of autoimmune and inflammatory disease, said method comprised of the steps of:

- 1) incubating an IKK α protein in the presence of a potential inhibitor of IKK α activity;
- 2) measuring the loss of IKK α activity;
- 25 3) comparing the amount of IKK α activity present in the absence of a potential inhibitor.

 Either the fully encoded IKK α or a fragment thereof can be used in the method for identifying a compound for the treatment of autoimmune and inflammatory disease.

 One particular advantage of using inhibitors specific to IKK α in the present
30 invention is that such inhibitors may have less side effects such as liver apoptosis as

compared to inhibitors for IKK β and NEMO. Other features and advantages of the invention will be apparent from the following detailed description of the invention.

BRIEF DESCRIPTION OF THE FIGURES

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Figure 1 shows the effect of IKK α and IKK β mRNA expression in HeLa cells after administration of siRNA directed to IKK α .

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Figure 2 shows siRNA directed to IKK α inhibits IKK α , IL-6 and IL-8 expression in HeLa cells.

Figure 3 shows IKK α siRNA inhibits NF- κ B activity in HeLa cells transfected with luciferase reporter gene.

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DETAILED DESCRIPTION OF THE INVENTION

I. General Description

The present invention provides a method for modulating NF- κ B dependent gene transcription, said method comprised of the step of modulating IKK α protein activity in a cell. The level of IKK α protein activity in a cell can be modulated upward or downward.

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The level of IKK α activity is preferentially modulated downward. One embodiment of the invention is based in part on the demonstration that the use of an IKK α specific inhibitor in TNF α stimulated human cells results in the modulation of genes under the influence of NF- κ B.

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The present invention employs siRNA for use in modulating the level of IKK α protein activity in the cell. SiRNA oligonucleotides directed to IKK α specifically hybridize nucleic acids encoding IKK α and interfere with IKK α gene expression. Accordingly, IKK α proteins levels are reduced and the total level of IKK α activity in the cell is reduced. Since IKK α has been shown to play a role in triggering the NF- κ B pathway (Table I; Li, X. et al, (2002) *J. Biol. Chem.*, 277: 45129-45140), which functions in the inflammatory response, compounds that have the property of being able to

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specifically and effectively inhibit IKK α are understood to be helpful in the treatment of autoimmune and inflammatory diseases.

Without intending to be limited by mechanism, it is believed that an IKK α specific
5 inhibitor acts by reducing the amount of activity of IKK α protein and or IKK α expression
in a cell, thereby directly or indirectly reducing the phosphorylation of NF- κ B p100
Pomerantz, J.L. and Baltimore, D., *Mol. Cell*, (2002) 10:693-5) or NF- κ B p65. Sizemore,
N. et al, *J. Biol. Chem.*, (2002) 277: 3863-9.

10 The present invention also provides methods for treating inflammatory and
autoimmune diseases using inhibitors of IKK α activity and is based in part on the
demonstration that the expression of proinflammatory genes under the influence of the NF-
 κ B pathway can be inhibited upon administering IKK α specific inhibitors to a cell. NF- κ B
dependent genes are found to be over expressed in autoimmune and inflammatory
15 conditions. Barnes et al, (1997) *New England J. Med.*, 336: 1066-1071; US
2002/0156000; US 6,395,545; 6,440,973; WO 02/060386 incorporated herein by reference.

The present invention also provides methods for identifying compounds that
modulate the activity of IKK α for the treatment of autoimmune and inflammatory disease.

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Definitions:

Unless defined otherwise, the scientific and technological terms and nomenclature
used herein have the same meaning as commonly understood by a person of ordinary skill
25 in the art to which this invention pertains.

Nucleotide sequences are presented herein by a single strand, in the 5' to 3'
direction, from left to right, using the one letter nucleotide symbols as commonly used in
the art and according with the recommendations of the IUPAC-IUB Biochemical
30 Nomenclature Commission (1972).

The term “**IKK α** ” as it is used herein refers to the alpha subunit of the I κ B kinase complex. IKK α is a kinase that phosphorylates I κ B, NF- κ B p100 or other protein substrates.

5 The term “**gene transcription**” as it is used herein means a process whereby one strand of a DNA molecule is used as a template for synthesis of a complementary RNA by RNA polymerase.

10 The term “**DNA**” as used herein refers to polynucleotide molecules, segments or sequences and is used herein to refer to a chain of nucleotides, each containing the sugar deoxyribose and one of the four adenine (A), guanine (G) thymine (T) or cytosine (C).

15 The term “**RNA**” as used herein refers to polynucleotide molecules, segments or sequences and is used herein to refer to a chain of nucleotides each containing the sugar ribose and one of the four adenine (A), guanine (G) uracil (U) or cytosine (C).

20 The term “**oligo**” as used herein means a short sequence of DNA or DNA derivatives typically 8 to 35 nucleotides in length. An oligonucleotide can be derived synthetically, by cloning or by amplification. The term “derivative” is intended to include any of the above described variants when comprising an additional chemical moiety not normally a part of these molecules. These chemical moieties can have varying purposes including, improving solubility, absorption, biological half life, decreasing toxicity and eliminating or decreasing undesirable side effects.

25 The term “**specifically hybridize**” as used herein means that under appropriate conditions a probe made or a nucleic acid sequence such as an siRNA oligo hybridizes, duplexes or binds only to a particular target DNA or RNA sequence present in a cell or preparation of DNA or RNA. A probe sequence such as an siRNA sequence specifically hybridizes to a target sequence when the base sequence of the probe nucleic acid and the
30 target sequence are complimentary to one another. The target sequence and the probe sequence do not have to be exactly complimentary to one another in order for the probe

sequence to specifically hybridize. It is understood that specific hybridization can occur when the target and probe sequences are not exactly complimentary to one another and specific hybridization can occur when up only about 80% of the bases are complimentary to one another. Preferably, it is understood that in specific hybridizations probe and target
5 sequence have 80% comprehensibility to one another. For discussions on hybridization see for example, Current Protocols in Molecular Biology, F. Ausubel et al., (ed.) Greene Publishing and Wiley-Interscience, New York (July, 2002).

The term "RNAi" as used herein means RNA interference process for a sequence-
10 specific post-transcriptional gene silencing or gene knockdown by providing a double-stranded RNA (dsRNA) that is homologous in sequence to the targeted gene. Small interfering RNAs (siRNAs) can be synthesized in vitro or generated by ribonuclease III cleavage from longer dsRNA and are the mediators of sequence-specific mRNA degradation.

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The term "**modulating IKK α activity**" as used herein means either inhibiting (decreasing) or stimulating (increasing) the level of activity of IKK α protein in a cell. IKK α activity can be modulated by modification of the levels and/or activity of IKK α protein, or by modification of the level of IKK α gene transcription and/or IKK 2 activity
20 structure such that the levels of IKK α protein activity in the cell is modulated. In the context of the present invention, inhibition is the preferred form of modulation.

The term "**autoimmune and inflammatory disease**" as used herein means diseases that are associated with autoimmune and inflammatory conditions such as
25 inflammatory and autoimmune conditions such as osteoarthritis, reperfusion injury, asthma, multiple sclerosis, Guillain-Barre syndrome, Crohn's disease, ulcerative colitis, psoriasis, graft versus host disease, systemic lupus erythematosus, rheumatoid arthritis, Alzheimer's disease, toxic shock syndrome, insulin-dependent diabetes mellitus, acute and chronic pain as well as symptoms of inflammation and cardiovascular disease, stroke,
30 myocardial infarction alone or following thrombolytic therapy, thermal injury, adult respiratory distress syndrome (ARDS), multiple organ injury secondary to trauma, acute

glomerulonephritis, dermatoses with acute inflammatory components, acute purulent meningitis or other central nervous system disorders, Grave's disease, myasthenia gravis, scleroderma and atopic dermatitis.

5 The term “**protein fragment**” as used herein means a truncated form of a protein. For IKK α suitable protein fragments should include the kinase domain understood as extending from at least amino acid residues 15 to 293 (SEQ. ID No. 2).

10 The term “**protein**” as used herein means isolated naturally occurring polypeptides, recombinantly produced proteins. Means for preparing such proteins are well understood in the art. Proteins may be in the form of the secreted protein, including truncated or mature forms. Proteins may optionally be modified to include an additional amino acid sequence which contains secretory or leader sequences, pro-sequences, sequences which aid in purification, such as multiple histidine residues, or an additional sequence for stability
15 during recombinant production. The proteins of the present invention are preferably provided in an isolated form, and preferably are substantially purified. A recombinantly produced version of a protein, including the secreted protein, can be substantially purified using techniques described herein or otherwise known in the art, such as, for example, by the one-step method described in Smith et al, *Gene*, 67:31-40 (1988). Proteins of the
20 invention also can be purified from natural, synthetic or recombinant sources using techniques described herein or otherwise known in the art.

 The term “**Gene knockdown**” as used herein refers to the reduction in the activity of a gene. The term “gene silencing” or gene inactivation are considered to have the same
25 meaning as the terms are used herein.

 The term “**potential inhibitor of IKK α activity**” as used herein means any compound or molecule that can cause the inhibition of IKK α activity. The inhibition of IKK α activity should be specific to IKK α . Potential IKK α inhibitors can be compounds
30 that block, antagonize, prevent, or reduce the activation of IKK α . IKK α inhibitors inhibit the activity of IKK α , preferably the *in vivo* activity, such that the catalytic activity of IKK α is inhibited 2) the phosphorylation of p100 or NF- κ B, p65 or I κ B is inhibited or 3) NF- κ B

dependent proinflammatory genes are inhibited. SiRNA as well as small molecule inhibitors directed to IKK α can be potential inhibitors of IKK α activity.

5 The term “**proinflammatory gene**” as used herein refers to any gene that is induced upon an inflammatory response through the NF- κ B pathway. Examples of proinflammatory genes include but are not limited to beta inhibin, IL-8, IL-6, interferon stimulated protein, TNF-induced protein, Cox2, GRO1 oncogene, CD44, interleukin 11, and superoxide dismutase.

10 The term “**specific inhibitor**” as used herein means an inhibitor that inhibits one protein more than another protein. For example, a potential inhibitor of IKK α is considered to be specific for IKK α over another IKK β protein when there is preferably at least 10 to 100 fold or greater and most preferably about 1000 fold difference in inhibition of IKK α compared to IKK β .

15 The term “**NF- κ B gene transcription**” as used herein means genes that are either upregulated or downregulated in response to the level of NF- κ B activity in a cell. Such genes include, but are not limited to IL-6, IL-8, inhibin, beta A, intercellular adhesion molecule 1, interferon stimulated protein, Cox2, IL-11, GRO1 and superoxide dismutase.

20 NF- κ B dependent genes are also discussed in US 2002/0156000: Barnes et al. (1997) *New England J. Med.* 336: 1066-1071; Pahl HL, *Oncogene*, (1999), 18, 6853-6866 incorporated herein by reference.

25 The term “**delivery**” as used herein refers to the introduction of foreign molecule (i.e. nucleic acid small molecule inhibitor) in cells.

 The term “**treating**” as used herein means the prevention, reduction, partial or complete alleviation or cure of a disease.

30 Using the present invention it is possible to observe the function of IKK α . In addition, specific siRNA oligos directed to IKK α have been designed and tested in human

cells showing a reduction in the expression of proinflammatory genes and NF- κ B target genes with their use. These siRNA and equivalent compounds may have therapeutic value in the treatment of autoimmune and inflammatory disease as described herein. It is therefore understood that compounds that inhibit either IKK α expression or IKK α protein activity also have therapeutic value.

The term “**Administration**” as used herein means the introduction of a Foreign molecule (i.e. nucleic acid, small molecule inhibitor) into a cell. The term is intended to be synonymous with the term “**Delivery**”.

III. Specific Embodiments

Preferred aspects of embodiments of the present invention are described in the following examples, which are not to be construed as limiting.

In one embodiment the method of the invention is used to reduce NF- κ B dependent gene expression in a cell, said method comprised of reducing IKK α protein activity in a cell. The level of NF- κ B dependent gene expression is proportionate to the level of IKK α activity in the cell. In other words, when the level of IKK α activity is lowered the level of expression of proinflammatory NF- κ B dependent genes is also reduced.

The method of the invention can comprise modulating NF- κ B dependent gene expression in a cell by administration of siRNA directed to IKK α . RNA interference is a method whereby siRNA can be used to knockdown or reduce the level of expression of a specific gene. In the case of IKK α , siRNA specifically directed to IKK α can be administered to cells in order to knockdown IKK α protein activity in the cell and to reduce the expression of NF- κ B proinflammatory genes. SiRNA can be designed according to the technique described by Tuschl, described as follows. Elbashir, SM et al, Nature, 2001, 411, 494-498. SiRNA that can efficiently knockdown a gene can be obtained by using siRNA duplexes composed of 21 nt sense and 21 nt antisense strands paired in a manner to have a 2-nt 3' overhang. The sequence of the 2-nt overhang is thought to make a

contribution to the specificity of the target recognition restricted to the unpaired nucleotide adjacent to the first base pair. 2-Deoxynucleotides are used in the 3' overhang.

5 The targeted region is selected from the human cDNA beginning at about 100 nt downstream of the start codon. Sequences can be searched for AA(N19)TT with approximately 40-60% G/C content. AA(N19) should match exactly the sequence of sense cDNA. The sequence of the sense siRNA corresponds to (N19)TT or N21, respectively. N19 exactly matches the sequence of sense cDNA. A blast search should be performed on the selected siRNA against genebank full-length genes and ESTs to ensure that only one
10 gene is targeted. The sequence of the siRNA should be selective to the target sequence.

Preparation of the siRNA duplexes

The siRNA duplexes used for delivery to cells can be prepared as follows. Approximately
15 0.02 to 0.2 μ M of the synthetic siRNAs can be used for delivery to various types of cells such as HeLa cells, Jurkat T cells, lymphocytes, HUVEC cells and fibroblasts. SiRNAs can be obtained from a number of sources including Dharmacon (Lafayette, CO) and Ambion (Austin, TX). The siRNA can be prepared by synthesizing the sense and antisense strand 21-nt oligos, followed by annealing of the single stranded oligos. The siRNA can be
20 incubated, pelleted and quantified using UV spectroscopy methods understood and used in the art.

Delivery of siRNA to cells and transfection of siRNA duplexes

25 Delivery of siRNA to cells can be performed according to cell transfection methods commonly used in the art. Elbashir SM et al, *Nature*, 2001, 411, 494-498; McManus MT et al, *J. Immunol.* 2002, 169:5754-60; Barton GM et al, *Proc. Natl. Acad. Sci.* (2002) 99:14943-5. Delivery of siRNA can be performed on various types of tissue culture cells. Preferably tissue culture cells of autoimmune or inflammatory significance such as
30 lymphocytes, epithelium cells and endothelial cells should be used. More specifically cells such as HeLa cells, Jurkat T cells, lymphocytes, HUVEC cells and fibroblasts. SiRNA can

be delivered to tissue and organisms as well. Lewis DL et al, *Nat. Genet.* (2002) 32:107-8; McCaffrey AP et al, *Nature* (2002) 418:38-39.

Various transfection reagents can be used for siRNA delivery such as lipids-
5 mediated transfection, electroporation or virus. In the preferred method the transfection reagent is OLIGOFECTAMINE™ available from Invitrogen (Carlsbad, CA). Transfection efficiencies should be between 40 and 100%.

For each sample between about 1 to 10 µg of siRNA duplex and about 100 µl of
10 Opti-MEM are mixed. In a separate tube 1 volume of Oligofectamine and 4 volumes of Opti-MEM are incubated from about 10 to 15 minutes at room temperature. The samples are then mixed and incubated for another 20 to 25 minutes at room temperature. Then 16 volumes of fresh Opti-MEM is added. SiRNA-transfection reagent is added to cultured cells (40 to 50% confluent). The cells are seeded for about 24 hours prior in antibiotics-
15 free medium using culture techniques commonly used in the art.

A knockdown effect should be found between 1 to 5 days after delivery of the siRNA. The amount of knockdown is generally 40 to 100 % of normal mRNA levels, and most preferably 60 to 100 % of normal mRNA levels.

20 Treatment of cells with a proinflammatory agent

In order to measure the extent of inhibition of NF-κB dependent proinflammatory genes, proinflammatory agents are administered to the cell. Acceptable proinflammatory
25 agents are compounds that induce expression of proinflammatory genes under the NF-κB pathway. Proinflammatory agents include but are not limited to TNFα, IL-1 and LPS. The preferred proinflammatory agent is TNFα. It is understood that other proinflammatory agents may effect expression of NF-κB dependent genes. The stimulation time and the amount of proinflammatory agent that is used will vary according to the agent used but
30 will be an amount sufficient to elicit a measurable proinflammatory response. TNFα is added to the cells with about 1 to 10 ng/ml for 30 minutes to 24 hours. Typically, the

proinflammatory agent is added before the measurement of proinflammatory genes is taken.

Preparation of RNA and PCR Primers

5

The level of gene knockdown or inhibition of gene transcription can be measured by analysis of mRNA from total RNA samples. Total RNA can be prepared between about 24 and 72 hrs after delivery siRNA using methods known to those skilled in the art. [www.invitrogen.com/transfection]. Preferably total cellular RNA is isolated from tissue or cell samples using the RNeasy™ kit and Rnase-Free DNase Set Protocol from Qiagen (Valencia, CA) according to the manufacturer's description.

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TaqMan Real-Time PCR Procedures

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PCR analysis can be used to analyze the isolated RNA and quantify the effects of the IKK α inhibitor on the transcription of NF- κ B dependent genes. PCR primers and/or probes used for the measurement of the transcription level of these genes can be prepared using techniques that are commonly used in the art. PCR primers should be designed for the amplification of the cDNA sequence from genes of interest. Software can be used to assist in designing design primers specific for target genes. Preferred software is Primer Express 1.5 Software (Applied Biosystems (Foster City, CA). Probes can be labeled with reporter agents such as fluorescent dye, FAM (6-carboxyfluorescein) at the 5' end and a fluorescent dye quencher TAMRA (6-carboxy-tetramethyl-rhodamine) at the 3' end. Other reporter agents commonly used in the art such as P³², S³⁵ fluorescein and Biotin can also be used. The specificity of PCR primers can be tested under normal PCR conditions in a thermal cycler prior to PCR quantitation. Total cellular RNA isolated from tissue or cell samples are used in reverse transcription (RT) reactions.

20

25

A "standard curve" can be constructed by plotting the C_t vs. the known copy numbers of the template in the standard. According to the standard curve, the copy numbers for all unknown samples are obtained automatically. To determine the copy numbers of the target transcript, a human genomic DNA (Clontech (Palo Alto, CA) can be used to generate a standard curve. The copy numbers of genomic DNA template are

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calculated according to the molecular weight of human diploid genome [3×10^9 bp = $3 \times 10^9 \times 660$ (M.W.) = 2×10^{12} g], and then 1 $\mu\text{g}/\mu\text{l}$ genomic DNA is converted into 2.4×10^6 copy numbers based upon the Avogadro's number (1 mol = 6.022×10^{23} molecules). Serial dilutions of the samples can be run in order to establish an estimate of the copy numbers. Copy numbers can be normalized to *GAPDH* or other housekeeping genes to minimize variability in the results due to differences in the RT efficiency and RNA integrity among test samples.

Microarray studies

Analysis of the transcription levels of genes can also be performed using microarray or cRNA chip analysis. These technologies allow the analysis of multiple genes in a single experiment. Preparation of cRNA, hybridization are performed according to methods as described herein and as otherwise commonly used in the art. Microarray analysis can be performed using procedures available from various companies such as Affymetrix and Agilent technologies.

The Affymetrix procedure is the preferred method and is performed essentially as follows: Between 5 and 10 micrograms of the total RNA can be converted into double stranded cDNA by reverse transcription using a cDNA synthesis kit. The preferred kit for cDNA synthesis is Superscript ChoiceTM, Invitrogen (Carlsbad, CA)) which has a special oligo (dT)24 primer (Genset, La Jolla, CA) containing a T7 RNA polymerase promoter site added 3' of the poly T tract. After second strand synthesis, labeled cRNA is generated from the cDNA samples by an *in vitro* transcription reaction using a reporting reagent such as biotin-11-CTP and biotin-16-UTP (Enzo, Farmingdale, NY). Labeled cRNA can be purified by techniques commonly used in the art. The preferred method is to use RNeasy spin columns (Qiagen, Valencia, CA). About 10 to 3 micrograms of each cRNA sample can be fragmented by mild alkaline treatment. Preferably, the cRNA sample is fragmented by treatment at 94° C for 35 minutes in fragmentation buffer as suggested by the manufacturer. A mixture of control cRNAs for bacterial and phage genes should be included to serve as tools for comparing hybridization efficiency between arrays and for relative quantitation of measured transcript levels. Before hybridization, the cRNA

samples are heated at about 94°C for 5 minutes, equilibrated at 45°C for 5 minutes and clarified by centrifugation (14,000 x g) at room temperature for 5 minutes. Aliquots of each cRNA sample are hybridized to arrays, according the manufacturer's directions. The arrays were then washed according to methods according to the manufacturer. The preferred wash is with non-stringent (6x SSPE, 0.01% Tween-20, 0.005% antifoam) and stringent (100 mM MES, 0.1M NaCl, 0.01% Tween 20), stained with R- Phycoerythrin Streptavidin- (Molecular Probes, Eugene, OR), washed again and scanned by an argon-ion laser scanner with the 560-nm long-pass filter (Molecular Dynamics; Affymetrix, Santa Clara, CA). Data analysis can be performed in order to determine if a gene expression level is increased, decreased or unchanged. Preferably, software such as MAS 5.0 software (Affymetrix, Santa Clara, CA) is used.

Identification of IKK α inhibitor compounds

Another embodiment of the invention provides a method for identifying a compound for the treatment of autoimmune and inflammatory disease, said method comprised of the steps of:

- 1) incubating an IKK α protein in the presence of a potential inhibitor of IKK α activity;
- 2) measuring the loss of IKK α activity;
- 3) comparing the amount of IKK α activity present in the absence of a potential inhibitor.

Compounds that are inhibitors of IKK α expression or IKK α activity for use in the treatment of autoimmune and inflammatory disease can be identified using the method of the invention. IKK α activity can be measured by determining the level of phosphorylation of the I κ B protein. Li, J. et al, 1998, *J. Biol. Chem.* 273:30736-41. A potential inhibitor of IKK α can be siRNA directed to IKK α or other small molecule inhibitor compounds that interact with the IKK α protein. Using the method of the invention siRNA directed to IKK α is delivered to cells according to the methods described herein or methods known in the art. In the case of siRNA inhibitors, the inhibitor compound does not have to be incubated with the IKK α protein. Compounds whose inhibitory activities are dependent on

interaction with IKK α protein should be incubated with the IKK α protein. It is understood that the IKK α siRNA does not have to be incubated with the IKK α protein in embodiments for identifying compounds for the treatment of autoimmune and inflammatory disease since IKK α siRNA inhibits IKK α protein activity without interacting or contacting the
5 IKK α protein. The method of the invention can be practiced using compounds that interact with IKK α protein and inhibit IKK α protein activity as well as using compounds that inhibit IKK α protein activity by inhibiting IKK α expression. After incubation and/or delivery of the potential inhibitor compound IKK α activity can be measured and then compared to the amount of IKK α activity present.

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This invention is further exemplified by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application, as well as the figures and sequence listing are hereby incorporated by reference. IKK α inhibitor compounds should be specific to
15 IKK α . Preferably IKK α inhibitor compounds are specific for IKK α over another IKK α protein when there is at least 10 to 100 fold or greater and most preferably about 100 fold difference in inhibition of the IKK α compared to IKK β .

20

The present invention also includes pharmaceutical compositions and formulations which include siRNA compounds as described herein. The pharmaceutical compositions can be administered topically, by inhalation, oral or parenteral as taught in US Patent No. 6,395,545, incorporated herein by reference. A preferred method of administration is as an emulsion or microemulsions. Another method of administration is through use of
25 liposomal formulations. Another method of administration using a "high pressure" delivery of RNAI into mammalian organs may also be used. See Nature Genetics Vol. 32 p107-108 incorporated herein by reference.

30

EXAMPLE 1: Preparation of siRNA duplexes.

SiRNA directed to IKK α sequence was designed according to the technique described by Tuschl. Elbashir SM et al, *Nature*, 2001, 411, 494-498. SiRNA duplexes composed of 21 nt sense and 21 nt antisense strands, paired in a manner to have a 2-nt 3' overhang were used. The sequence of the 2-nt overhang is thought to make a contribution to the specificity of the target recognition restricted to the unpaired nucleotide adjacent to the first base pair. 2-Deoxynucleotides were used in the 3' overhang.

The targeted region was selected from the cDNA for human IKK α shown in SEQ. ID No. 1 beginning 100 nt downstream of the start codon. Sequences were searched for AA(N19)TT and with approximately 50% G/C content. The sequence of the sense siRNA corresponded to (N19)TT to N21, respectively. The 3' end of the sense RNA was converted to TT. A blast search was performed on the selected siRNA against EST libraries to ensure that only one gene was targeted. The sequences selected are shown as SEQ. ID Nos. 2 – 7.

Approximately 0.2 micromoles of the synthetic siRNAs were obtained from Dharmacon Research Inc. (Lafayette, Colorado). The siRNAs were desalted and deprotected by the supplier and therefore were not further gel purified. The siRNA oligos were annealed and shipped in 4 tubes. Each tube was added 1 ml sterile RNase-free water to make 20 μ M siRNA concentrations. After 1 to 2 hours of incubation on ice the siRNAs were ready for use in transfection.

Example 2: Delivery of siRNA to HeLa cells

Transfection of siRNA duplexes

Delivery of siRNA duplexes was performed with OLIGOFECTAMINETM reagent available from Invitrogen (Carlsbad, CA). The samples were prepared in a 6 well format. Transfection efficiencies were found to be about 80%.

For each well of a 6 well plate, one tube containing 10 μ l of 20 μ M siRNA duplex with 90 μ l of Opti-MEM, and in a separate tube 4 μ l of OLOGOFECTAMINE™ reagent with 96 μ l of Opti-MEM were mixed and incubated for 7-10 minutes at room temperature. The two tubes were combined and incubated for another 20 to 25 minutes at room temperature. Then 800 μ l of fresh Opti-MEM was added to obtain a final solution of 1000 μ l. Then 1000 μ l of siRNA-OLIGOFECTAMINE™ was added to cultured cells (40 to 50% confluent). The cells were seeded the previous day in 6- well plates at a density of 2×10^5 cells/well using 2 ml of DMEM tissue culture medium supplemented with 10% FBS without antibiotics. The control used for transfection was inverted siRNA. A knockdown effect was generally found after 1-2 days.

Example 3: Preparation of RNA and PCR Primers

Total RNA was prepared from the cells 2 days after delivery of siRNA's. Total cellular RNA was isolated from tissue or cell samples using the RNeasy™ kit and RNase-Free DNase Set Protocol from Qiagen (Valencia, CA) according to the manufacturer's directions. PCR primers and TaqMan probes were designed using Primer Express 1.5 Software (Applied Biosystems, Foster, CA). The sequence of the PCR primers used were SEQ. ID. No. 9: 5'-GCACAGAGATGGTGAAAATCATTG-3', and SEQ. ID. No. 10: 5'-CAACTTGCTCAAATGACCAAACAG-3'. The probe sequence SeQ. ID No. 11: 5'-TGAGCACACGGTCCTGACTCTGCA labeled with a reporter fluorescent dye, FAM (6-carboxyfluorescein), at the 5' end and a fluorescent dye quencher TAMRA (6-carboxy-tetramethyl-rhodamine) at the 3' end. The specificity of PCR primers was tested under normal PCR conditions in a thermal cycler prior to TaqMan™ PCR quantitation. Total cellular RNA was isolated from cell samples using the RNeasy Kits and RNase-Free DNase Set Protocol according to the manufacturer's description (Qiagen). Reverse transcription (RT) reactions were carried out for each RNA sample in MicroAmp reaction tubes using TaqMan™ reverse transcription reagents. Each reaction tube contained 500 ng of total RNA in a volume of 50 μ l containing $1 \times$ TaqMan™ RT buffer, 5.5 mM $MgCl_2$, 500 μ M of each dNTP, 2.5 μ M of Random Hexamers or oligo-d(T)₁₆ primers, 0.4 U/ μ l of RNase inhibitor, and 1.25 U/ μ l of MultiScribe Reverse Transcriptase. RT reactions were

carried out at 25°C for 10 min, 48°C for 40 min and 95°C for 5 min. Real-time PCR was performed in a MicroAmp Optical 96-Well Reaction Plate (Applied Biosystems). Each well contained 2 µl of each RT product (20 ng total RNA), 1× TaqMan buffer A, 5.5 mM MgCl₂, 200 µM dATP/dCTP/dGTP, 400 µM dUTP, 200 nM primers (forward and
5 reverse), 100 nM TaqManTM probe, 0.01 U/µl AmpErase, and 0.025 U/µl AmpliTaqTM Gold DNA polymerase in a total volume of 25 µl. Each well was closed with MicroAmp Optical caps (Applied Biosystems), following complete loading of reagents.

Amplification conditions were 2 min at 50°C (for AmpErase UNG incubation to remove any uracil incorporated into the cDNA), 10 min at 95°C (for AmpliTaqTM Gold activation),
10 and then run for 40 cycles at 95°C for 15 s, 60°C for 1 min. All reactions were performed in the ABI Prism 7700 Sequence Detection System for the test samples, standards, and no template controls. They were run in triplicates using the Sequence Detector V 1.6 program. The R_n and C_t were averaged from the values obtained in each reaction. A "standard curve" was constructed by plotting the C_t vs. the known copy numbers of the
15 template in the standard. According to the standard curve, the copy numbers for all unknown samples were obtained automatically. To determine the copy numbers of the target transcript, a human genomic DNA (Clontech, Palo Alto, CA) was used to generate a standard curve. The copy numbers of genomic DNA template were calculated according to the molecular weight of human diploid genome [3×10^9 bp = $3 \times 10^9 \times 660$ (M.W.) = 2
20 $\times 10^{12}$ g], and then 1 µg/µl genomic DNA was converted into 2.4×10^6 copy numbers based upon the Avogadro's number (1 mol = 6.022×10^{23} molecules). The genomic DNA was serially (every ten-fold) diluted at a range of 5×10^5 to 5×10^0 copy numbers. Each sample was run in triplicates, and the R_n (the ratio of the amount of reporter dye emission to the quenching dye emission) and threshold cycle (C_t) values were averaged from each
25 reaction. The copy numbers were then normalized to *GAPDH* to minimize variability in the results due to differences in the RT efficiency and RNA integrity among test samples.

Example 4: Microarray studies using Affymetrix

30 Preparation of cRNA and gene chip analysis.

Preparation of cRNA, hybridization and scanning of the U133A were performed according to the manufacturer's protocol (Affymetrix, Santa Clara, CA). Briefly, 5-10 µg of the total RNA was converted into double stranded cDNA by reverse transcription using a cDNA synthesis kit (Superscript Choice™, Invitrogen Carlsbad, CA with a special oligo(dT024 primer) (Genset, La Jolla, CA) containing a T7 RNA polymerase promoter site added 3' of the poly T tract. After second strand synthesis, labeled cRNA was generated from the cDNA samples by an *in vitro* transcription reaction supplemented with biotin-11-CTP and biotin-16-UTP (Enzo, Farmingdale, NY). The labeled cRNA was purified by using RNeasy spin columns (Qiagen, Valencia, CA). Fifteen micrograms of each cRNA sample was fragmented by mild alkaline treatment at 94°C for 35 minutes in fragmentation buffer (40 mM Tris-acetate, pH 8.1, 100 mM potassium acetate, 30 mM magnesium acetate) and then used to prepare 0.3 ml of master hybridization mix (100 mM MES, 1M [NaCl], 20mM EDTA, 0.01% Tween 20, 0.1 mg/ml herring sperm DNA (Promega, Madison, WI), 0.5 mg/ml acetylated BSA (Invitrogen Carlsbad, CA). A mixture of control cRNAs for bacterial and phage genes available from the manufacturer was included in the mix (BioB, BioC, BioD, and cre, at 1.5, 5, 25 and 100 pM, respectively) to serve as tools for comparing hybridization efficiency between arrays and for relative quantitation of measured transcript levels. Before hybridization, the cRNA samples were heated at 94°C for 5 minutes, equilibrated at 45 ° C for 5 minutes and clarified by centrifugation (14,000 x g) at room temperature for 5 min. Aliquots of each sample (10 µg of cRNA in 200 µl of the master mix) were hybridized to U133A arrays (Affymetrix) at 45 ° C for 16 h in a rotisserie oven set at 60 rpm. The arrays were then washed with non-stringent (6x SSPE, 0.01% Tween-20, 0.005% antifoam) and stringent (100 mM MES, 0.1M NaCl, 0.01% Tween 20), stained with R- Phycoerythrin Streptavidin- (Molecular Probes, Eugene, OR), washed again and scanned by an argon-ion laser scanner with the 560-nm long-pass filter (Molecular Dynamics; Affymetrix). Data analysis was performed by using MAS5.0 software (Affymetrix, Santa Clara, CA). The software includes algorithms that determine whether a gene is absent or present (absolute call) and whether the expression level of a gene in an experimental sample is significantly increased or decreased (difference call) relative to a control sample. To assess differences in gene expression, Affymetrix used the "Signal Log Ratio Algorithm" which calculates

signal log ratio values using a one-step Tukey's Biweight method by taking a mean of the log ratios of probe pair intensities across the two arrays. Fold change values were calculated using the following formula: $2^{\text{Signal Log Ratio}}$, if Signal Log Ratio ≥ 0 ; $(-1) \cdot 2^{-(\text{Signal Log Ratio})}$, if Signal Log Ratio < 0 (Affymetrix GeneChip Expression Analysis manual).

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Example 5: Inhibition of IKK α gene expression by siRNA directed to IKK α in HeLa cells.

Figure 1 demonstrates that siRNA of IKK α inhibits mRNA expression of IKK α in HeLa cells. HeLa cells were transfected with IKK α _3 (SEQ. ID. NO. 2) or IKK α _3vt
10 inverted control of oligos of IKK_3 (SEQ. ID. No. 3). by OLIGOFECTAMINE™ using the methods described herein. IKK α mRNA copy numbers were determined by TaqMan™. A 3 fold reduction was found in samples receiving the siRNA relative to samples that had received the control.

15

Example 6: IKK α SiRNA does not inhibit IKK β expression.

IKK α _3 siRNA specifically inhibited the mRNA expression of IKK α but had no effect on IKK β mRNA expression as shown in Figure 1 and Table I. This is expected since the sequence of IKK α siRNA was designed to specifically target IKK α mRNA but
20 not the IKK β mRNA. Thus, IKK α _3 siRNA is demonstrated to be a specific IKK α inhibitor with no crossing activity towards IKK β .

Example 7: Inhibition of IKK α expression modulated the expression of other genes in the NF- κ B pathway.

25

Table 1 shows selected IKK α -regulated genes listed in order of their mRNA fold changes in 2 hr TNF α stimulated HeLa cells treated with IKK α siRNA (IKK α _3) vs. 2 hr TNF α stimulated HeLa cells treated with inverted control siRNA. Data were obtained by DNA microarray studies using Affymetrix U133A chips. Results from two independent
30 chip hybridizations are shown. The mRNA of IKK β and house keeping genes such as beta Actin and GAPDH were not affected by IKK α siRNA (NC, no change call by Affymetrix MAS 5.0 analysis). Inhibition of IKK α expression by IKK α -3 siRNA decreased the

expression of known NF- κ B-dependent genes such as IL-6, IL-8, IL-11, Cox-2, Dihydrodiol dehydrogenase 1, TNF-induced proteinIntegrin, Urokinase-type plasminogen activator receptor; Intercellular adhesion molecule 1 (CD54), Bone morphogenetic protein 2, Interferon-stimulated protein, 15 kDa, Superoxide dismutase 2 and GRO1. These genes
5 are understood to be NF- κ B targets. Pahl HL, *Oncogene* (1999)18, 6853-6866; Li X et al, 2002, J. Biol. Chem., 277: 45129-45140.

TABLE 1

Accession	Gene name	F Id changes (IKK α siRNA+TNF_vs_ctrl siRNA+TNF)		TNF stimulation
		Exp.1	Exp.2	
M13436	Inhibin, beta A	-10.6	-7.4	Yes
NM_002133	Heme oxygenase (decycling) 1	-5.5	-5.9	No
AF043337	Interleukin 8	-2.5	-2.3	Yes
NM_001353	Dihydrodiol dehydrogenase 1	-2.3	-1.9	NO
NM_014350	TNF-induced protein	-2.2	-1.8	Yes
NM_002205	Integrin, alpha 5	-2.0	-2.3	No
AY029180	Urokinase-type plasminogen activator receptor	-1.9	-1.8	No
NM_000201	Intercellular adhesion molecule 1 (ICAM1/CD54)	-1.7	-2.0	Yes
M24915	CD44 antigen	-1.6	-1.7	Yes
AA583044	Bone morphogenetic protein 2	-1.6	-1.4	yes
NM_000600	Interleukin 6	-1.5	-1.5	Yes
NM_000963	Cox2	-1.5	-1.7	Yes
NM_000641	Interleukin 11	-1.5	-1.6	Yes
NM_005101	Interferon-stimulated protein, 15 kDa	-1.5	-1.6	Yes
W46388	Superoxide dismutase 2	-1.4	-1.8	Yes
NM_001511	GRO1 oncogene	-1.4	-1.5	yes
M33197	GAPDH	NC	NC	No
X00351	Actin, beta	NC	NC	No
AF080157	IKK α	-3.1	-2.8	No
AF080158	IKK β	NC	NC	No

NF- κ B activity is modulated by inhibition of IKK α as shown in NF- κ B reporter assay system.

Example 8:

The inhibition of IKK α expression modulated the activity of NF- κ B in NF- κ B-dependent reporter assay. Figure 3 shows that TNF α -induced NF- κ B-dependent
5 luciferase activity was inhibited by siRNA of IKK α . The pNF- κ B-Luc plasmid contains NF- κ B promoter elements which can be activated by NF- κ B. Downstream of the promoter is the luciferase reporter gene. When activation of NF- κ B is induced by TNF α , NF- κ B will activate the artificial promoter and the promoter will drive the luciferase reporter gene expression. In this experiment, we used HeLa cells that had been stably transfected with
10 the pNF- κ B-Luc reporter (Stratagene: 219078, La Jolla, CA).

Cells were transfected with siRNA as follows:

6 μ g of siRNA duplex and 100 μ l of Opti-MEM were mixed. In a separate tube 6 μ l Oligofectamine and 24 μ l Opti-MEM were incubated for 10 minutes at room
15 temperature. The samples were then mixed and incubated for 25 minutes at room temperature before 64 μ l of fresh Opti-MEM was added to obtain a final solution of 200 μ l. Finally, 200 μ l of siRNA-transfection reagent was added to cultured cells seeded 24 hours prior to the transfection in antibiotics-free medium. Two days later cells were stimulated with TNF α (10 ng/ml) for 24 hours. Cells were then harvested and counted. 5 X
20 10^5 cells in 100 μ l of PBS were added to wells of 96-well plate and the luciferase activity was measured by LucLite Luciferase Reporter Gene Assay Kit (Packard BioScience, The Netherlands). The luciferase activity was induced by TNF α (comparing IKK α _3inverted+TNF vs. IKK α _3inverted_unstim, Fig. 3) due to activation of NF- κ B. However, such induction was significantly reduced (~ 3 fold reduction) by IKK α _3 siRNA
25 (comparing IKK α _3+TNF vs. IKK α _3_unstim, Fig. 3). Thus IKK α _3 siRNA blocked TNF α -induced NF- κ B activity in gene transcription. The data demonstrate that IKK α plays an important role in TNF α -induced NF- κ B-mediated gene transcription.

Example 9: siRNA of IKK α inhibits IL-6 and IL-8 expression in HeLa cells

Figure 2 show that siRNA of IKK α inhibits the expression of proinflammatory genes under the influence of the genes in the NF- κ B pathway including IL-6 and IL-8 in HeLa cells. HeLa cells were transfected according to the methods described herein. An approximately 2 fold reduction in the expression of IL-6 was found in TNF α stimulated cells. In un-stimulated cells there was approximately 2 fold reduction in IL-6 mRNA. A 2.6 fold reduction in the level of IL-8 mRNA in TNF α stimulated HeLa cells was found and approximately 2.6 fold reduction was found IL-8 mRNA in unstimulated HeLa cells. These data show the correlation between the level of IKK α activity and the level of expression of genes that are under the influence of the NF- κ B pathway.

The level of IL-6 and IL-8 have been shown to be elevated in autoimmune and inflammatory disease conditions Ishihara K et al. *Cytokine Growth Factor Rev* (2002),13:357-68; Mukaida N, *Int J Hematol* (2000),72:391-8]. Monoclonal antibody of IL-6 has been used for treatment for systemic lupus erythematosus [<http://www.clinicaltrials.gov/>]. A fully human anti-IL8 monoclonal antibody (ABX-IL8) has also been applied to patients with chronic bronchitis and COPD [<http://www.clinicaltrials.gov/>]. Therefore, the treatment of inflammatory disease by inhibitors IL-6-IL-8 activity is demonstrated as a method for testing representative autoimmune and inflammatory diseases.

In addition to IL-6 and IL-8 which are important targets for inflammatory diseases and autoimmune diseases (see above), many other IKK α -regulated genes (Table I) are also mediators of inflammatory diseases and autoimmune diseases.

Cyclooxygenase 2 (Cox2), also known as prostaglandin endoperoxide synthase 2, is the key enzyme required for the conversion of arachidonic acid to prostaglandins which regulate immunity and inflammation (Harris SG et al (2002) *Trends in Immunology*. 23:144-50; Turini ME and DuBois RN. (2002) *Annual Review of Medicine*. 53:35-57). Cox2 mediates inflammation and selective Cox2 inhibitors have been shown to be potent

antiinflammatory agents (Rodrigues CR. et al, 2002, *Current Medicinal Chemistry*. 9:849-67).

5 Inhibin, beta A (Activin A) is a member of the transforming growth factor beta (TGF-beta) superfamily and functions in inflammatory pathways. Jones KL. et al (2000) *Endocrinology* 141:1905-8; Yu, J. et al (1997) *Cytokines Cellular & Molecular Therapy*. 3:169-77. There is correlation between inhibin secretion and damage of seminiferous tubules in a model of experimental autoimmune orchitis (Suescun MO. et al, 2001, *Journal of Endocrinology* 170:113-20).

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Adhesion molecules such as /intercellular adhesion molecule-1 (ICAM1), integrin alpha 5, CD44 and Gro1 are important players in autoimmunity and inflammatory responses. Long-term reversal of established autoimmunity was observed upon the transient blockade of the LFA-ICAM pathway. Bertry-Coussot, L. et al (2002) *Journal of Immunology* 168:3641-8. Blockade of the ICAM1 and LFA-1 interaction is an effective approach for immunosuppression. Yusuf-Makagiansar H. et al (2002) *Medicinal Research Reviews* 22:146-67. Integrin alpha 5 is one component of the alpha 5 beta 1 integrin which mediates integrin-independent neutrophil recruitment to endotoxin-induced lung inflammation. Burns JA. et al (2001) *Journal of Immunology* 166:4644-4649. Alpha 5 beta 1 integrin activates an NF-κB-dependent program of gene expression important for angiogenesis and inflammation. Klein S et al (2002) *Molecular & Cellular Biology* 22:5912-22. CD44, a cell-adhesion molecule that is ubiquitously expressed on leukocytes and parenchymal cells, functions in several inflammatory diseases. Pure E. et al (2001) *Trends in Molecular Medicine* 7:213-21. Functional activation of lymphocyte CD44 in peripheral blood is a marker of autoimmune disease activity. Estess P. et al (1998) *Journal of Clinical Investigation* 102:1173-82. Antibodies to CD44 prevent central nervous system inflammation and experimental encephalomyelitis by blocking secondary leukocyte recruitment. Brocke S. et al (1999) *Proceedings of the National Academy of Sciences of the United States of America* 96:6896-901.

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